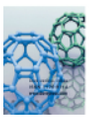
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International Journal of Nanomedicine



ISSN: (Print) (Online) Journal homepage: www.tandfonline.com/journals/dijn20

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To cite this article: Krupa Kansara, Pal Patel, Ritesh K Shukla, Alok Pandya, Rishi Shanker, Ashutosh Kumar & Alok Dhawan (2018) Synthesis of biocompatible iron oxide nanoparticles as a drug delivery vehicle, International Journal of Nanomedicine, 13:sup1, 79-82, DOI: 10.2147/IJN.S124708

To link to this article: https://doi.org/10.2147/IJN.S124708

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Published online: 15 Mar 2018.

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International Journal of Nanomedicine

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SHORT REPORT

Synthesis of biocompatible iron oxide nanoparticles as a drug delivery vehicle

This article was published in the following Dove Press journal:
International Journal of Nanomedicine

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Abstract: Over the last decade, there has been growing interest in developing novel nanoparticles (NPs) for biomedical applications. A safe-by-design approach was used in this study to synthesize biocompatible iron oxide NPs. The size of the particles obtained was ~100 nm. Although these NPs were significantly ($P<0.05$) internalized in MCF-7 (human breast adenocarcinoma cell line) cells, no adverse effect was observed in the cells as assessed by cytotoxicity assays (neutral red uptake and 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) and cell cycle analysis. Our data demonstrate the potential of iron oxide NPs as a biocompatible carrier for targeted drug delivery.

Keywords: biocompatible iron oxide nanoparticles, human breast adenocarcinoma cells, coprecipitation

Introduction

Iron-based magnetic nanoparticles (NPs) such as magnetite (Fe_3O_4) have been studied in detail due to their unique properties, such as stability over time, biocompatibility, and sensitivity to magnetic field.[1–3] They can potentially be used as magnetic targeted drug delivery carriers and magnetic resonance imaging contrast agents due to their high saturation magnetization, low toxicity, and biocompatibility.[4] The magnetic properties of Fe_3O_4 NPs are attributed to their size and size distribution, which, in turn, is dependent on the route of synthesis.

Therefore, in this study, an attempt was made to synthesize Fe_3O_4 NPs using a safe-by-design approach by the coprecipitation method. Polyethylene glycol (PEG) was used as surfactant to control the particle size and narrow size distribution. The biocompatibility of Fe_3O_4 NPs was evaluated by cytotoxicity assays and cell cycle analysis in the human breast adenocarcinoma cell line (MCF-7).

Materials and methods

Materials

Ferric chloride hexahydrate and ferrous sulfate were purchased from SD-Fine-Chem. Ltd, Mumbai, India. Ployetheleneglycol (PEG-6000), dimethylesulphoxide, sodium hydroxide (NaOH), minimum essential medium eagle, phosphate-buffered saline, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), and antibiotic-antimycotic solution were purchased from HiMedia Laboratories Pvt. Ltd., (Mumbai, India). The MCF-7 cell line was purchased from the National Centre for Cell Sciences, Pune, India.

Fe_3O_4 NP synthesis

The preparation of Fe_3O_4 NPs was performed by a chemical coprecipitation method of Fe^{2+} and Fe^{3+} ions (1:2 molar ratios) by the addition of NaOH.[5] A total volume of 15 mL

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submit your manuscript | www.dovepress.com
Dovepress
http://dx.doi.org/10.2147/IJN.S124708

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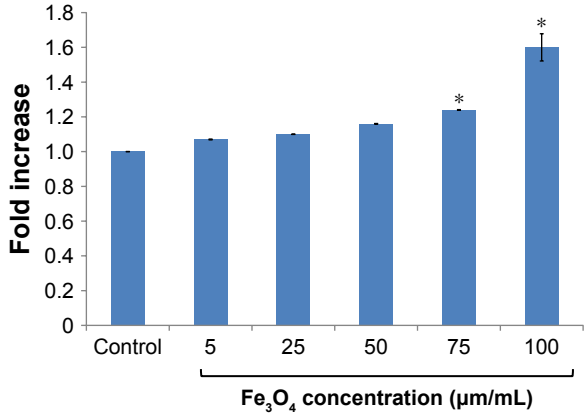


Figure 1 Internalization of Fe_3O_4 NPs in MCF-7 cells using flow cytometry.
Notes: Data are expressed as mean ± standard error of the mean from three independent experiments. *$P<0.05$, when compared with control.
Abbreviations: NPs, nanoparticles; MCF-7, human breast adenocarcinoma cell line.

of 0.25 M Fe^{2+} and 0.5 M Fe^{3+} solutions were prepared in deionized water. PEG was then added and the temperature slowly risen up to 80°C. During the initial 2 minutes of the reaction, NaOH was added to achieve a pH of 10. The reaction was allowed to continue on a magnetic stirrer for 2 hours. Thereafter, the suspension was centrifuged and washed several times with deionized water to lower the pH to 7. Finally, the particles were suspended in 10 mL of dimethylesulphoxide.

Characterization of Fe_3O_4 NPs

One milliliter of the stock suspension of Fe_3O_4 NPs was diluted in 10 mL complete minimum essential medium eagle. The hydrodynamic size and zeta potential were determined using Zetasizer Nano ZS.

Cytotoxicity assessment

The cytotoxic potential of Fe_3O_4 NPs was assessed in MCF-7 cells after 6 and 24 hours of treatment using MTT and neutral red uptake (NRU) assays as described by Mosmann[6] and Borenfreund and Puerner, respectively.[7]

Cellular internalization of NPs

The internalization of Fe_3O_4 NPs in MCF-7 cells was assessed according to the method described in our earlier study.[8]

Cell cycle analysis

The effect of Fe_3O_4 NPs on cell cycle was assessed according to the method described in our earlier study.[8]

Results and discussion

The mean hydrodynamic size and zeta potential of synthesized Fe_3O_4 NPs were 98.19±1.0 nm and 36±2 mV, respectively. Flow cytometric analysis revealed a significant ($P<0.05$) increase in the internalization of Fe_3O_4 NPs in MCF-7 cells after 24 hours exposure at the two higher concentrations, as evident by an increase in the side scatter intensity (Figure 1).

In the cytotoxicity assays, namely NRU and MTT, Fe_3O_4 NPs were found to be biocompatible as there was no significant increase in the NRU (88% at concentration 150 μM/mL) and a reduction (96%) in the mitochondrial succinate dehydrogenase activity was observed at the highest concentration after 24-hour exposure (Figures 2A and B).

Additionally, no change in cell cycle progression was observed in Fe_3O_4 NPs-treated MCF-7 cells, after 24-hour exposure (Figure 3).

Conclusion

Our results demonstrated that Fe_3O_4 NPs synthesized using the safe-by-design approach showed no adverse effect on cells, as assessed by cytotoxicity assays and cell cycle analysis in MCF-7 cells, even though they are significantly

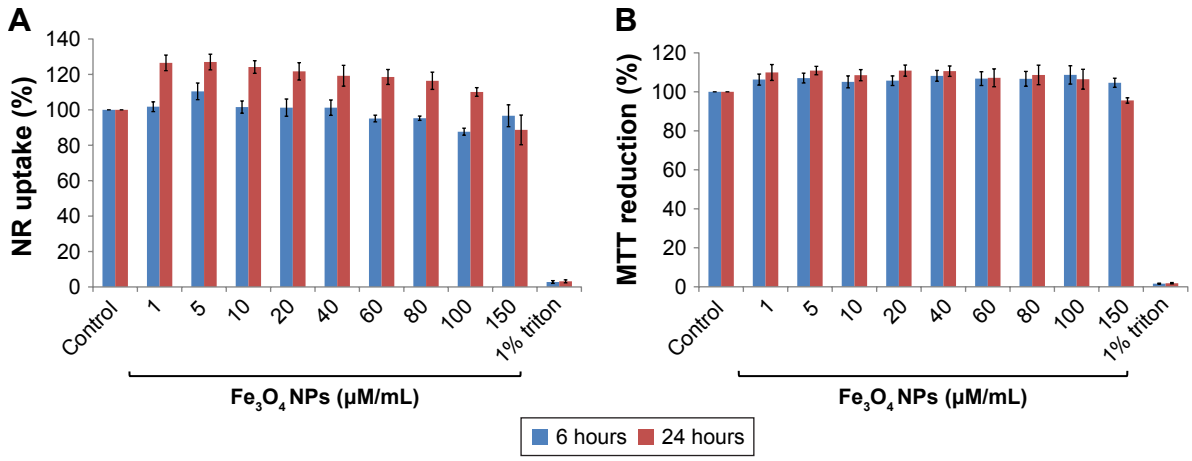


Figure 2 Cytotoxicity of Fe_3O_4 NPs in MCF-7 cells. (**A**) NR uptake (%); (**B**) MTT reduction (%).
Notes: The viability of the control cells was considered as 100%. Data are expressed as mean ± standard error of the mean from three independent experiments.
Abbreviations: NPs, nanoparticles; MCF-7, human breast adenocarcinoma cell line; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NR, neutral red.

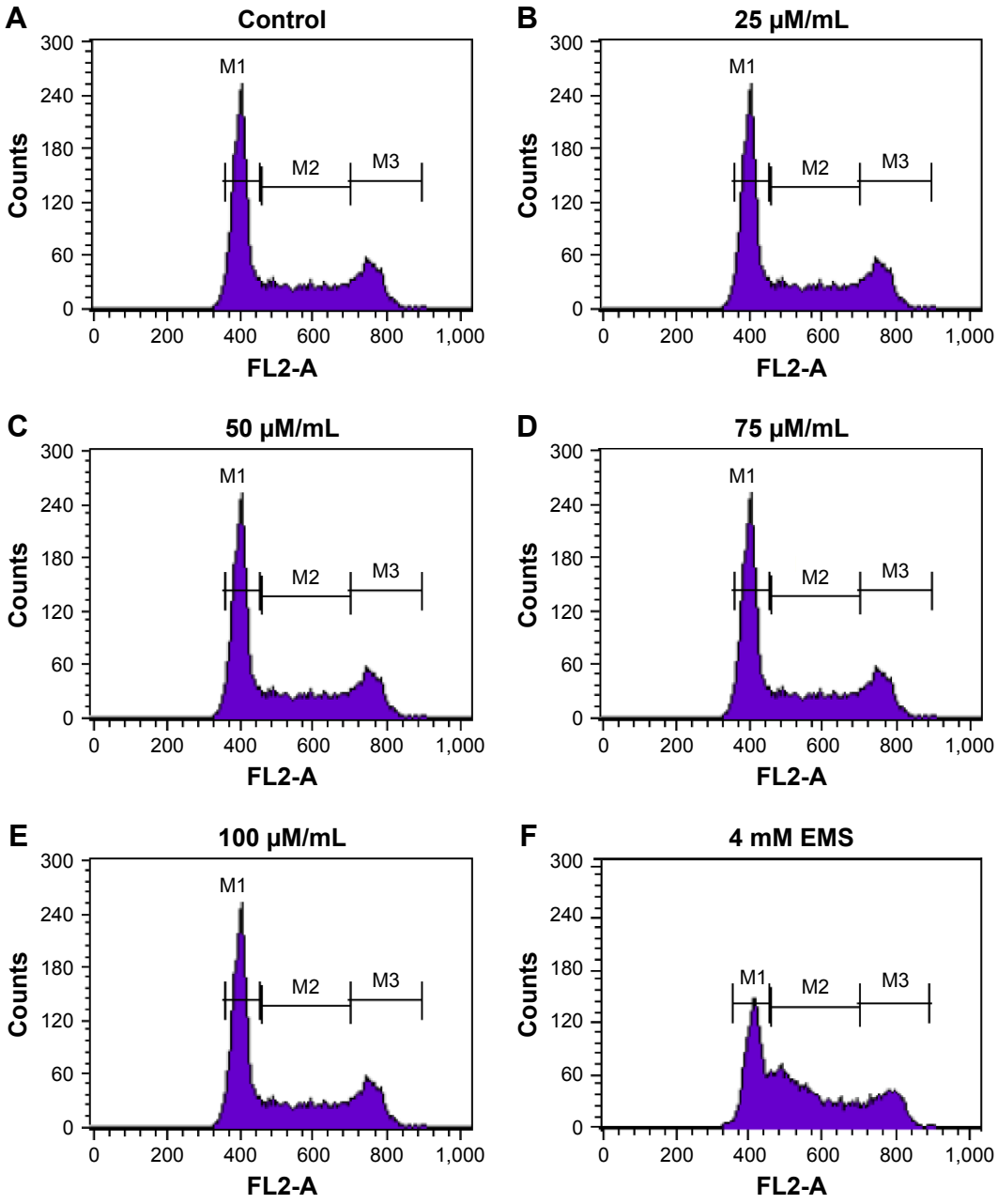


Figure 3 Effect of Fe_3O_4 NPs on cell cycle progression in MCF-7 cells (**A**) control (**B**–**E**) cells treated with different concentration of Fe_3O_4 NPs and (**F**) cells treated with 4 mM EMS.

Abbreviations: EMS, ethylmethane sulfonate; NPs, nanoparticles; MCF-7, human breast adenocarcinoma cell line.

internalized. Therefore, these NPs have a potential to be used as a carrier for targeted drug delivery.

Acknowledgments

The authors acknowledge the funding from the Centre for Nanotechnology Research and Application (CENTRA) by The Gujarat Institute for Chemical Technology (GICT); and the European Union Seventh Framework Programme (FP7/2007–2013) under grant agreement number 263147 (NanoValid – Development of reference methods for hazard identification, risk assessment and LCA of engineered nanomaterials).

Disclosure

The authors report no conflicts of interest in this work.

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